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Short Communication

Research Progress of Sirt1 in Lung Injury

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1. Abstract

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2. Keywords

Sirt1; lung injury; COPD; lung fibrosis; FOXOs; oxidative stress; NF-κB

Recently lung injury has a high clinical morbidity and mortality due to the destroyed alveolar epithelial barrier and endothelial dysfunction. If there is no further treatment, it will be deteriorated into pulmonary fibrosis and even lung cancer. Silencing regulator 2 related enzyme 1 (Sirt1) is a nicotinamide adenine dinucleotide (NAD+)-dependent deacetylase, associated with various cellular processes, such as proliferation, apoptosis and inflammation, through deacetylated histones and some non-histone proteins, such as the fork head transcription factor (FOXO) and nuclear factor κB (NF-κB), et al. Sirt1 plays a key role in smoking-induced lung injury, bronchial asthma, and oxidative stress-induced lung disease. It has been found that resveratrol, as a strongest agonist of polyphenols Sirt1, plays an obvious anti-inflammatory effect by de acetylation of several inflammatory related transcription factors, such as NF-κB. Therefore, Sirt1 is a negative regulatory target of inflammatory factors, and Sirt1 signaling pathways can provide a new direction for the treatment of lung injury. In this review, we will discuss about the research progress of Sirt1 in lung injury.

3. Introduction

Sirtuins is a family of Histone De Acetylase (HDACs) family, which catalyzes the de acetylation of histone and non- histone lysine residues [1]. The human Sirt1 gene is located in the chromosome 10q21.3 and gene length is 33.72kb, which mainly located in the nucleus and expressed in the eukaryotes and prokaryotes. The mammalian Sirtuins family has seven members (Sirt1-Sirt7), which share a common highly conserved catalytic domain with different N- and C-terminal structures, and different terminal extensions determine their subcellular localization and distribution [2], distributed in mitochondria, cytoplasm, nucleus and tissues. Sirt1, 6 and 7 are mainly located in the nucleus, where Sirt1 and Sirt2 can be located in the nucleus or in the cytoplasm [3,4]. Sirt3, 4 and 5 are mainly located in mitochondria, mainly involved in stress response, aging, cardiac metabolic diseases and liver metabolic pathways [5-8]. In addition, Sirtl also has a certain expression in the cytoplasm, and there is a nuclear-cytoplasmic shuttle movement to adjust the balance and homeostasis of all aspects in vivo. Studies have shown that Sirt1-3 has strong deacetylase activity, while SIRT4-7 has weaker deacetylase activity [9]. Sirt1 is involved in chromatin modification, epigenetics, inflammation, and stress responses, interacting with histones and some non-histone substrates. Among them, non-histone substrates include that tumor suppressor gene p53, nuclear factor-κB (NF-κB), peroxisome proliferator-activated receptor γ co-activator 1α (PGC-1α), FOXO, liver x (LXR), PARP, Ku70 and hypoxia-inducible factor (HIF)-1α, and so on [10]. Moreover, Sirt1 regulates metabolism by de acetylation of PGC-1α, which plays an important role in glucose and lipid metabolism during fasting, and is involved in regulating glucose metabolism, lipid metabolism, cell differentiation, cell stress, apoptosis and inflammatory reactions and so on. Sirt1 regulates glucose metabolism because FOXO1, as a de acetylating transcription factor and co-activating factor peroxisome proliferator, activates receptor gamma co-activator 1α (PGC1α), and enhances the process of sugar isogenesis. Meanwhile, Sirt1 is also expressed in degenerative diseases, acute myeloid leukemia, colon cancer, prostate cancer, ovarian cancer, breast cancer, melanoma and lung adenocarcinoma; Sirt2 is mainly expressed in neurodegenerative diseases, brain tissue and glioma; Sirt3 is mainly expressed in neurodegenerative diseases, B-cell chronic lymphocytic leukemia, mantle cell lymphoma, chronic lymphocytic leukemia, breast

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cancer and gastric cancer; Sirt4 and Sirt6 are mainly expressed in breast cancer and colon cancer Sirt5 is mainly expressed in pancreatic cancer, breast cancer and non-small cell carcinoma; Sirt7 is mainly expressed in cancer: liver, testis, spleen, thyroid and breast [11]. Studies have shown that rs12413112 and rs1467568 polymorphisms have a protective effect on cardiovascular diseases, regarding the location of this polymorphism in the 3′UTR, and many studies have revealed an association between the nucleotide variations in this position and change in the expression pattern of their flanking genes. Hence, it seems that the A-allele of this polymorphism increases Sirt1 expression that results in reduced risk of developing cardiovascular diseases. A significant association between the rs1467568 polymorphism and the risk of type II diabetes during a fatal confrontation with famine, hence Sirt1 is a crucial genetic factor that effects fetal programming during malnutrition.

Alveolar epithelial cells are the first protective barrier, which are also an important respiratory barrier for gas exchange. When inhaled toxic gases and particles by cigarette smoke, epithelial cells cause inflammation and further form a feedback loop for chronic inflammation [12,13]. Pulmonary inflammatory response is a feature of many lung diseases, such as Chronic Obstructive Pulmonary Disease (COPD) [14]. It is widely studied in clinical and scientific research as well as in the mechanism of lung injury. In clinical practice, lung injury occurs mainly due to bacteria or viral infections, inhalation syndrome and severe shock [15]. Inflammation activates alveolar macrophages, which in turn produce cytokines and inflammatory chemokines, which produce reactive oxygen species and reactive nitrogen-induced pulmonary edema [16,17]. Severe pneumonia causes the production of cytokines and chemokines, as well as polymorphonuclear leukocyte infiltration and reactive oxygen species, which in turn leads to damage to the epithelium and endothelium of the lungs, and leads to increased alveolar permeability and hinder to alveolar gas exchange [18]. Changes in the permeability of the alveolar epithelium and endothelial barrier are the pathophysiological features of lung injury, and the pulmonary endothelium plays a key role in lung injury [19,20]. Once activated, endothelial cells secrete cytokines and inflammatory chemokines, such as tumor necrosis factor TNF-α, interleukin IL-1β, and MCP-1 cause the accumulation of neutrophils and macrophages, which in turn leads to dysfunction of cells and organs. The recovery of lung injury requires a complete endothelial barrier and reconstruction as well as alveolar epithelial barrier repair. Sirt1 inhibits the process of inflammation by regulating a variety of mediators, which expression increase can inhibit the release of inflammatory factors, such as the release of tumor necrosis factor-α and interleukin (IL)-8. Therefore, Sirt1 becomes a process that inhibits the development of inflammation and potential targets [21].

3.1. The role of Sirt1 in Chronic Obstructive Pulmonary Disease (COPD)

COPD is one of the global public health challenges at present. Smoking induces oxidative stress, endoplasmic reticulum stress and apoptosis [22], which is the main cause of Chronic Obstructive Pulmonary Disease (COPD) and lung cancer [23]. COPD refers to chronic airflow limitations that cause permanent alveolar wall loss and alveolar space enlargement, thus the deletion of alveolar junction Structural and alveolar space enlargement are the most important pathological changes in COPD. Sirt1 is a transcriptional regulator that inhibits the expression of pro-apoptotic proteins [24], which can de acetylation a variety of transcription factors, for example, it can de acetylation p^{53} , causing p^{53} homeostasis imbalance and reducing the expression of Bax in the downstream gene [25], and then inhibits apoptosis. If the down regulation of Sirt1 expression can increase the level of $p⁵³$ acetylation, and then promote apoptosis and senescence [26,27]. Studies have shown that Sirtl is a key factor in regulating oxidative stress in organisms, and can de acetylation of non -histone proteins such as antioxidants in cells. If the expression is suppressed, oxidative stress in cells will increase, which accelerate cell aging and damage oxidative stress to accelerate the aging process, and Sirt1 can inhibit oxidative stress, endothelial dysfunction and inflammatory response, and also participate in the development of COPD. It is reported that Sirt1 is a long-lived factor, which involve in the development of COPD through regulating oxidative stress, inflammation and senescence [28]. The latest research shows that Sirt1 protects the oxidative stress induced by smoking and de acetylation of FOXO3 [29]. Because FOXO3 is the substrate of Sirt1 nonhistone, Sirtl can regulate the expression of antioxidant genes and enzymes in vivo by de acetylation of FOX03, thus reducing the damage of oxidative stresses products to cells and delaying cell apoptosis and aging. Because Sirt1 can de acetylate FOXOs, p53 and NF-κB, which can control the development process of COPD [30]. The up-regulated expression of Sirt1 reduces the level of acetylation of FOXO3 and protects the acute lung injury by regulating the downstream of antioxidant and anti-apoptotic factors [31-34]. The histopathological changes in acute lung injury include the following aspects: (1) the accumulation of neutrophils and macrophages after being activated; (2) apoptosis; (3) the increase of pro inflammatory and pulmonary capillary of permeability [35-37]. The expression of Sirt1 in the lung tissue of smokers was reduced in the nucleus [38], and the expression of Sirt1 was significantly reduced in smokers and COPD patients. In clinical, monitoring the level of Sirtl in the lung tissue of patients may provide a new way of thinking for the treatment of COPD.

3.2. The role of Sirt1 in lung fibrosis

Fibrosis is usually associated with aging and disease, all over the body and the formation of excessive fibrous connective tissue, which is caused by typical organ damage and eventually leads to organ failure [39]. The development process of fibrosis is usually divided into four main stages [40]. The first stage is primarily injury of organs and tissues that is the injury of epithelial cells and main cells; the second stage is the activation of the fibrous effector cells; the third stage is the deposition of the extracellular matrix; the fourth stage is the promotion of the dynamic deposition of extracellular matrix. The development of fibrosis eventually leads to organ failure and even death [25,41]. At the early stage of lung injury, lung tissue is surrounded by chronic inflammation, and subsequently the outer tissue wall is damaged by collagen [1,42]. Different pathological findings show that accumulation of extracellular matrix components is a common feature of fibrosis [7,43]. The abnormal accumulation of extracellular matrix, which is a typical fibrosis feature, leading to damage of tissue structure and function of [43,44]. Clinical chemical poisoning causes high morbidity and mortality worldwide. It is reported that there are more than two million cases, dying of parquet poisoning each year in China [45-48], in which parquet poisoning death is due to acute lung injury and multiple organ failure [49], however, parquet poisoning is the main cause of lung injury caused by oxygen free radicals [50,51]. As an inflammatory disease, lung injury can leading to alveolar damage, aeration perfusion to varying degrees, and non-cardiogenic pulmonary edema, and eventually leads to death [52]. There are various causes of fibrosis, including acute lung injury, chronic inflammation, autoimmune reaction and genetic changes.

Clinically, chemical poisoning causes high morbidity and mortality worldwide. It is reported that China has died of parquet poisoning more than two million cases each year^[45-48], among which parquet poisoning death is due to acute lung injury and more Organ failure^[49], lung injury caused by parquet poisoning is mainly the production of oxygen free radicals [50,51]. As an inflammatory disease, lung injury lead to alveolar damage, varying degrees of ventilatory perfusion, and non-cardiogenic pulmonary edema, and ultimately death [52]. The FOXO family belongs to the fork head transcription factor family and is an important cell fate regulator [53], which transfers from cytoplasm to nucleus and plays a transcriptional role by binding to promoter of target gene. It has been shown that acetylation of FOXO increases nuclear transcription and promotes transcriptional activity [54], and FOXO can be deacetylated by Sirt1 to enhance binding and transcriptional activity with DNA [55]. Therefore, Sirt1 can inhibit the development of pulmonary fibrosis through de acetylation of FOXO, which may provide a new strategy for the treatment of pulmonary fibrosis.

3.3. The role of Sirt1 in lung cancer

Among all cancers, lung cancer is one of the highest morbidity and mortality worldwide [56], of which approximately 80% are Non-Small Cell Lung Cancer (NSCLC) [57]. The role of Sirt1 in the mechanism of tumorigenesis and development has been paid more and more attention. Sirt1 may play a role in inhibiting cancer and promoting cancer in cancer. Recently, Sirtl is closely related to the metastasis of tumor cells, which can be used as a target for the treatment of lung cancer patients and its expression can also be one of the prognostic indicators of NSCLC treatment. Sirt1 and AMPK are closely related to the development of non-small cell lung cancer [58,59]. AMPK widely exists in organisms and plays a key role in cell energy metabolism and cell growth and differentiation. It is widely believed that AMPK has great potential in the treatment of cardiovascular diseases, diabetes, obesity, central nervous system diseases and cancer [60,61]. However, by H&E staining, Western blot, RT-PCR and immunofluorescence test, it was proved that the expression of Sirt1 and AMPK in NSCLC decreased significantly compared with in normal group [62]. Further exploration of the role of Sirt1 and AMPK in NSCLC provides a new direction for the treatment of NSCLC. Sirt1 may be a new target for the treatment of NSCLC, which brings new ideas for the clinical targeting therapy of NSCLC and the research and development of related targeted drugs. Furthermore, Sirtl plays an important role in the occurrence, development, treatment and prognosis of lung cancer, but its accurate mechanism is still not clear, which still needs further study. Whether Sirt1 has been used as an accelerator or inhibitor in the process of tumor formation and development has yet to be conclusive. However, it is believed that with the continuous discovery of the molecules interacting with SIRT1 and the deepening of the research on the correlation between SIRT1 and tumor, the mechanism of SIRT1 in the formation of tumor may be a new target for the treatment of tumor.

4. Conclusions

In short, the increase of the pro inflammatory mediators leads to lung injury [63]. It is reported that IL-6 is the medium for inducing acute lung injury and pulmonary edema, and Sirt1 plays a key role in regulating the development of inflammation, and it plays an anti-inflammatory effect by controlling the release of pro-inflammatory factors [64,65]. Oxidative stress is considered to be an important factor in cell damage. Oxygen free radicals can cause membrane lipid peroxidation and protein oxidation to destroy cell integrity and induce apoptosis. Sirtl is a deacetylase dependent on NAD⁺, which plays a role in antioxidant stress, anti-inflammatory, and inhibition of apoptosis through the role of histone / non histone de acetylation group. A large number of studies have found that Sirtl has a strong anti-oxidative stress in neurodegenerative diseases, cardiovascular system diseases and renal system

diseases, which mechanism is still unclear, and further discussion is needed. Sirt1 plays a key role in the death of apoptotic cells, cell senescence and endothelial growth in the stress response pathway [66], but the mechanism of protecting blood vessels from stress induced endothelial dysfunction should be further explored. However, there are not many researches on Sirt1 in lung injury, so further exploration is needed. Research on the treatment strategies of Sirt1 agonists for lung injury and other diseases will be the direction of future research, and actively explore the pathogenesis of Sirtl in the regulation of respiratory diseases and the accurate target of its role, and actively develop its agonists and inhibitors in the clinical appliance of the broad prospects.

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References

1. Winnik S, Auwerx J, Sinclair DA, Matter CM. Protective effects of sirtuins in cardiovascular diseases: from bench to bedside. Eur Heart J. 2015;36(48):3404-12.

2. Mei Z, Zhang X, Yi J, Huang J, He J, Tao Y. Sirtuins in metabolism, DNA repair and cancer. J Exp Clin Cancer Res. 2016;35(1):182.

3. Lin J, Sun B, Jiang C, Hong H, Zheng Y. Sirt2 suppresses inflammatory responses in collagen-induced arthritis. Biochem Biophys Res Commun. 2013;441(4):897-903.

4. Haigis MC, Guarente LP. Mammalian sirtuins--emerging roles in physiology, aging, and calorie restriction. Genes Dev. 2006;20(21):2913- 21.

5. He W, Newman JC, Wang MZ, Ho L, Verdin E. Mitochondrial sirtuins: regulators of protein acylation and metabolism. Trends Endocrinol Metab. 2012;23(9):467-76.

6. van de Ven RAH, Santos D, Haigis MC. Mitochondrial Sirtuins and Molecular Mechanisms of Aging. Trends Mol Med. 2017;23(4):320-31.

7. Sack MN, Finkel T. Mitochondrial metabolism, sirtuins, and aging. Cold Spring Harb Perspect Biol. 2012;4(12).

8. Parihar P, Solanki I, Mansuri ML, Parihar MS. Mitochondrial sirtuins: emerging roles in metabolic regulations, energy homeostasis and diseases. Exp Gerontol. 2015;61:130-41.

9. Ma Q, Wood TK. Protein acetylation in prokaryotes increases stress resistance. Biochem Biophys Res Commun. 2011;410(4):846-51.

10. Nakagawa T, Guarente L. Sirtuins at a glance. J Cell Sci. 2011;124(Pt 6):833-8.

11. Carafa V, Rotili D, Forgione M, Cuomo F, Serretiello E, Hailu GS, et al. Sirtuin functions and modulation: from chemistry to the clinic. Clin Epigenetics. 2016;8:61.

12. Pace E, Ferraro M, Siena L, Scafidi V, Gerbino S, Di Vincenzo S, et al. Carbocysteine regulates innate immune responses and senescence processes in cigarette smoke stimulated bronchial epithelial cells. Toxicol Lett. 2013;223(2):198-204.

13. Pouwels SD, Heijink IH, ten HNH, Vandenabeele P, Krysko DV, Nawijn MC, et al. DAMPs activating innate and adaptive immune responses in COPD. Mucosal Immunol. 2014;7(2):215-26.

14. Lee H, Abston E, Zhang D, Rai A, Jin Y. Extracellular Vesicle: An Emerging Mediator of Intercellular Crosstalk in Lung Inflammation and Injury. Front Immunol. 2018;9:924.

15. Bhattacharya J, Matthay MA. Regulation and repair of the alveolarcapillary barrier in acute lung injury. Annu Rev Physiol. 2013;75:593- 615.

16. Hoogerwerf JJ, de Vos AF, Levi M, Bresser P, van der Zee JS, Draing C, et al. Activation of coagulation and inhibition of fibrinolysis in the human lung on bronchial instillation of lipoteichoic acid and lipopolysaccharide. Crit Care Med. 2009;37(2):619-25.

17. van Zoelen MA, Verstege MI, Draing C, de Beer R, van't Veer C, Florquin S, et al. Endogenous MCP-1 promotes lung inflammation induced by LPS and LTA. Mol Immunol. 2011;48(12-13):1468-76.

18. Fanelli V, Ranieri VM. Mechanisms and clinical consequences of acute lung injury. Ann Am Thorac Soc. 2015;12 Suppl 1:S3-8.

19. Maniatis NA, Orfanos SE. The endothelium in acute lung injury/ acute respiratory distress syndrome. Curr Opin Crit Care. 2008;14(1):22- 30.

20. Matthay MA, Zemans RL. The acute respiratory distress syndrome: pathogenesis and treatment. Annu Rev Pathol. 2011;6:147-63.

21. Howitz KT, Bitterman KJ, Cohen HY, Lamming DW, Lavu S, Wood JG, et al. Small molecule activators of sirtuins extend Saccharomyces cerevisiae lifespan. Nature. 2003;425(6954):191-6.

22. Zhang L, Guo X, Xie W, Li Y, Ma M, Yuan T, et al. Resveratrol exerts an anti-apoptotic effect on human bronchial epithelial cells undergoing cigarette smoke exposure. Mol Med Rep. 2015;11(3):1752-8.

23. Kato R, Mizuno S, Kadowaki M, Shiozaki K, Akai M, Nakagawa K, et al. Sirt1 expression is associated with CD31 expression in blood cells from patients with chronic obstructive pulmonary disease. Respir Res. 2016;17(1):139.

24. Matsushita N, Takami Y, Kimura M, Tachiiri S, Ishiai M, Nakayama

T, et al. Role of NAD-dependent deacetylases SIRT1 and SIRT2 in radiation and cisplatin-induced cell death in vertebrate cells. Genes Cells. 2005;10(4):321-32.

25. Shi X, Wei W, Wang N. Tremella polysaccharides inhibit cellular apoptosis and autophagy induced by Pseudomonas aeruginosa lipopolysaccharide in A549 cells through sirtuin 1 activation. Oncol Lett. 2018;15(6):9609-16.

26. Luo J, Li M, Tang Y, Laszkowska M, Roeder RG, Gu W. Acetylation of p53 augments its site-specific DNA binding both in vitro and in vivo. Proc Natl Acad Sci U S A. 2004;101(8):2259-64.

27. Vaziri H, Dessain SK, Ng EE, Imai SI, Frye RA, Pandita TK, et al. hSIR2(SIRT1) functions as an NAD-dependent p53 deacetylase. Cell. 2001;107(2):149-59.

28. Taka C, Hayashi R, Shimokawa K, Tokui K, Okazawa S, Kambara K, et al. SIRT1 and FOXO1 mRNA expression in PBMC correlates to physical activity in COPD patients. Int J Chron Obstruct Pulmon Dis. 2017;12:3237-44.

29. Yao H, Sundar IK, Ahmad T, Lerner C, Gerloff J, Friedman AE, et al. SIRT1 protects against cigarette smoke-induced lung oxidative stress via a FOXO3-dependent mechanism. Am J Physiol Lung Cell Mol Physiol. 2014;306(9): L816-28.

30. Lavu S, Boss O, Elliott PJ, Lambert PD. Sirtuins--novel therapeutic targets to treat age-associated diseases. Nat Rev Drug Discov. 2008;7(10):841-53.

31. Chai D, Zhang L, Xi S, Cheng Y, Jiang H, Hu R. Nrf2 Activation Induced by Sirt1 Ameliorates Acute Lung Injury After Intestinal Ischemia/ Reperfusion Through NOX4-Mediated Gene Regulation. Cell Physiol Biochem. 2018;46(2):781-92.

32. Gu L, Tao X, Xu Y, Han X, Qi Y, Xu L, et al. Dioscin alleviates BDLand DMN-induced hepatic fibrosis via Sirt1/Nrf2-mediated inhibition of p38 MAPK pathway. Toxicol Appl Pharmacol. 2016;292:19-29.

33. Zhou Z, You Z. Mesenchymal Stem Cells Alleviate LPS-Induced Acute Lung Injury in Mice by MiR-142a-5p-Controlled Pulmonary Endothelial Cell Autophagy. Cell Physiol Biochem. 2016;38(1):258-66.

34. Ding YW, Zhao GJ, Li XL, Hong GL, Li MF, Qiu QM, et al. SIRT1 exerts protective effects against paraquat-induced injury in mouse type II alveolar epithelial cells by deacetylating NRF2 in vitro. Int J Mol Med. 2016;37(4):1049-58.

35. Rudiger A, Singer M. Mechanisms of sepsis-induced cardiac dysfunction. Crit Care Med. 2007;35(6):1599-608.

36. Matuschak GM, Lechner AJ. Acute lung injury and the acute respiratory distress syndrome: pathophysiology and treatment. Mo Med.

2010;107(4):252-8.

37. Grommes J, Soehnlein O. Contribution of neutrophils to acute lung injury. Mol Med. 2011;17(3-4):293-307.

38. Yang SR, Wright J, Bauter M, Seweryniak K, Kode A, Rahman I. Sirtuin regulates cigarette smoke-induced proinflammatory mediator release via RelA/p65 NF-kappaB in macrophages in vitro and in rat lungs in vivo: implications for chronic inflammation and aging. Am J Physiol Lung Cell Mol Physiol. 2007;292(2):L567-76.

39. Xin Z, Ma Z, Hu W, Jiang S, Yang Z, Li T, et al. FOXO1/3: Potential suppressors of fibrosis. Ageing Res Rev. 2018;41:42-52.

40. Rockey DC, Bell PD, Hill JA. Fibrosis--a common pathway to organ injury and failure. N Engl J Med. 2015;372(12):1138-49.

41. Martin K, Pritchett J, Llewellyn J, Mullan AF, Athwal VS, Dobie R, et al. PAK proteins and YAP-1 signalling downstream of integrin beta-1 in myofibroblasts promote liver fibrosis. Nat Commun. 2016;7:12502.

42. Wang YY, Qiu XG, Ren HL. Inhibition of acute lung injury by rubriflordilactone in LPS-induced rat model through suppression of inflammatory factor expression. Int J Clin Exp Pathol. 2015;8(12):15954-9.

43. Baues M, Dasgupta A, Ehling J, Prakash J, Boor P, Tacke F, et al. Fibrosis imaging: Current concepts and future directions. Adv Drug Deliv Rev. 2017;121:9-26.

44. Murray LA. Editorial: The Cell Types of Fibrosis. Front Pharmacol. 2015;6:311.

45. Hao R, Li P, Wang Y, Wu Z, Song H. Chemical poisoning-related injury in China. Lancet. 2013;382(9901):1327-8.

46. Onyeama HP, Oehme FW. A literature review of paraquat toxicity. Vet Hum Toxicol. 1984;26(6):494-502.

47. Sabzghabaee AM, Eizadi-Mood N, Montazeri K, Yaraghi A, Golabi M. Fatality in paraquat poisoning. Singapore Med J. 2010;51(6):496-500.

48. Gunnell D, Eddleston M, Phillips MR, Konradsen F. The global distribution of fatal pesticide self-poisoning: systematic review. BMC Public Health. 2007;7:357.

49. Sittipunt C. Paraquat poisoning. Respir Care. 2005;50(3):383-5.

50. Blanco-Ayala T, Andérica-Romero AC, Pedraza-Chaverri J. New insights into antioxidant strategies against paraquat toxicity. Free Radic Res. 2014;48(6):623-40.

51. Sun S, Jiang Y, Wang R, Liu C, Liu X, Song N, et al. Treatment of Paraquat-Induced Lung Injury With an Anti-C5a Antibody: Potential Clinical Application. Crit Care Med. 2018;46(5):e419-25.

52. Ding Q, Liu GQ, Zeng YY, Zhu JJ, Liu ZY, Zhang X, et al. Role of

IL-17 in LPS-induced acute lung injury: an in vivo study. Oncotarget. 2017;8(55):93704-11.

53. Kaletsky R, Lakhina V, Arey R, Williams A, Landis J, Ashraf J, et al. The C. elegans adult neuronal IIS/FOXO transcriptome reveals adult phenotype regulators. Nature. 2016;529(7584):92-6.

54. Sewastianik T, Szydlowski M, Jablonska E, Bialopiotrowicz E, Kiliszek P, Gorniak P, et al. FOXO1 is a TXN- and p300-dependent sensor and effector of oxidative stress in diffuse large B-cell lymphomas characterized by increased oxidative metabolism. Oncogene. 2016;35(46):5989-6000.

55. Wang B, Yang Q, Sun YY, Xing YF, Wang YB, Lu XT, et al. Resveratrol-enhanced autophagic flux ameliorates myocardial oxidative stress injury in diabetic mice. J Cell Mol Med. 2014;18(8):1599-611.

56. Subramaniam S, Thakur RK, Yadav VK, Nanda R, Chowdhury S, Agrawal A. Lung cancer biomarkers: State of the art. J Carcinog. 2013;12:3.

57. Tsim S, O'Dowd CA, Milroy R, Davidson S. Staging of non-small cell lung cancer (NSCLC): a review. Respir Med. 2010;104(12):1767-74.

58. Chua KF, Mostoslavsky R, Lombard DB, Pang WW, Saito S, Franco S, et al. Mammalian SIRT1 limits replicative life span in response to chronic genotoxic stress. Cell Metab. 2005;2(1):67-76.

59. Zhang YJ, Xiang H, Liu JS, Li D, Fang ZY, Zhang H. Study on the mechanism of AMPK signaling pathway and its effect on apoptosis of human hepatocellular carcinoma SMMC-7721 cells by curcumin. Eur Rev Med Pharmacol Sci. 2017;21(5):1144-50.

60. Funai K, Cartee GD. Inhibition of contraction-stimulated AMPactivated protein kinase inhibits contraction-stimulated increases in PAS-TBC1D1 and glucose transport without altering PAS-AS160 in rat skeletal muscle. Diabetes. 2009;58(5):1096-104.

61. Gaidhu MP, Fediuc S, Anthony NM, So M, Mirpourian M, Perry RL, et al. Prolonged AICAR-induced AMP-kinase activation promotes energy dissipation in white adipocytes: novel mechanisms integrating HSL and ATGL. J Lipid Res. 2009;50(4):704-15.

62. Yang F. The expression and mechanism of Sirt1 and AMPK in nonsmall cell lung cancer. J BUON. 2018;23(1):106-10.

63. Lan KC, Chao SC, Wu HY, Chia-Lien Chiang, Ching-Chia Wang, Shing-Hwa Liu, et al. Salidroside ameliorates sepsis-induced acute lung injury and mortality via downregulating NF-κB and HMGB1 pathways through the upregulation of SIRT1. Sci Rep. 2017;7(1):12026.

64. Kauppinen A, Suuronen T, Ojala J, Kaarniranta K, Salminen A. Antagonistic crosstalk between NF-κB and SIRT1 in the regulation of inflammation and metabolic disorders. Cell Signal. 2013;25(10):1939- 48.

65. Yeung F, Hoberg JE, Ramsey CS, Keller MD, Jones DR, Frye RA, et al. Modulation of NF-kappaB-dependent transcription and cell survival by the SIRT1 deacetylase. EMBO J. 2004;23(12):2369-80.

66. Michan S, Sinclair D. Sirtuins in mammals: insights into their biological function. Biochem J. 2007;404(1):1-13.